

# **Quantifying variation in female internal genitalia: no evidence for plasticity in response to sexual conflict risk in a seed beetle**

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1 **Abstract**

2 Sexually antagonistic coevolution can drive the evolution of male traits that harm females,  
3 and female resistance to those traits. While males have been found to vary their harmfulness  
4 to females in response to social cues, plasticity in female resistance traits remains to be  
5 examined. Here we ask whether female seed beetles *Callosobruchus maculatus* are capable  
6 of adjusting their resistance to male harm in response to the social environment. Among seed  
7 beetles, male genital spines harm females during copulation and females might resist male  
8 harm via thickening of the reproductive tract walls. We develop a novel Micro-CT imaging  
9 technique to quantify female reproductive tract thickness in 3-dimensional space, and  
10 compared the reproductive tracts of females from populations that had evolved under high  
11 and low levels of sexual conflict, and for females reared under a social environment that  
12 predicted either high or low levels of sexual conflict. We find little evidence to suggest that  
13 females can adjust the thickness of their reproductive tracts in response to the social  
14 environment. Neither did evolutionary history affect reproductive tract thickness.  
15 Nevertheless, our novel methodology was capable of quantifying fine-scale differences in the  
16 internal reproductive tracts of individual females, and will allow future investigations into the  
17 internal organs of insects and other animals.

18 **Key Words:** female genital evolution, sexual conflict, sexually antagonistic coevolution,  
19 phenotypic plasticity, experimental evolution, sex ratio

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21

## 22 **1. Introduction**

23 Evolutionary conflict between the sexes can arise over the expression of traits that improve  
24 the fitness of one sex but are detrimental to the fitness of their sexual partners (Arnqvist and  
25 Rowe 2005; Parker 2006). Sexual selection favours traits in males that confer greater  
26 fertilisation success regardless of the outcomes for females, while antagonistic selection on  
27 females can favour morphological and behavioural traits that function to resist harm induced  
28 by males (Stockley 1997; Edvardsson and Tregenza 2005). Harmful traits in males are  
29 diverse in form and include: male harassment during courtship and mating (den Hollander  
30 and Gwynne 2009; Rankin et al. 2011), infanticide (Arnqvist and Rowe 2005; Parker 2006),  
31 and toxic ejaculates (Chapman et al. 1995; den Hollander and Gwynne 2009). One  
32 conspicuous form of sexual conflict is the damage imposed to the female's reproductive tract  
33 by male genitalia during copulation (Lange et al. 2013). Genital damage during copulation is  
34 found across a variety of taxa, particularly within arthropod lineages (Crudginton and Siva-  
35 Jothy 2000; Blanckenhorn et al. 2002; Kamimura 2010; Tataric and Cassis 2010; Kamimura  
36 2012; Tataric et al. 2014) and is considered to be a by-product of male traits enhancing  
37 fertilisation success, rather than harm directly benefitting males (Morrow et al. 2003; Hotzy  
38 and Arnqvist 2009; Grieshop and Polak 2014; McNamara et al. 2020).

39 The evolution of harmful traits in one sex is expected to generate selection on the  
40 opposite sex favouring the evolution of traits to resist harm, initiating sexually antagonistic  
41 coevolution between the sexes (Arnqvist and Rowe 1995; Rice 1996; Arnqvist and Rowe  
42 2002a; Tataric and Cassis 2010; Dougherty et al. 2017). While comparative evidence  
43 suggests that coevolution of harmful male traits and female resistance traits is widespread  
44 (Brennan et al. 2007; Perry and Rowe 2012; Hopwood et al. 2016; Dougherty et al. 2017),  
45 whether these traits can respond plastically to an individual's environment has received less  
46 attention. Males of a wide range of taxa have been found to adjust ejaculate size, composition

47 and quality in response to exposure to rival males (Gage and Baker 1991; Pilastro et al. 2002;  
48 Evans et al. 2003; Simmons et al. 2007; Thomas and Simmons 2007; DelBarco-Trillo 2011),  
49 and recent work suggests that male genital morphology can be adjusted in response to the  
50 competitive environment experienced during sexual development (Brennan et al. 2017;  
51 André et al. 2018). Thus, males exhibit phenotypic plasticity in sexual traits that can be costly  
52 for females. Firman and Simmons (Firman and Simmons 2013) found that female house mice  
53 exposed to greater levels of sperm competition risk produced ova with lower fertilizability,  
54 presumably to mitigate against the threat of polyspermy when males compete for  
55 fertilizations. This finding suggests that females too may be capable of responding to the  
56 socio-sexual environment, and reduce the costs associated with male competition. Using a  
57 model species in which sexual conflict has been widely documented, we test the hypothesis  
58 that sexual conflict favours the evolution of phenotypic plasticity in female resistance traits in  
59 response to the immediate risk of sexual conflict.

60         Seed beetles (*Callosobruchus maculatus*) are a widely used model species for sexual  
61 conflict studies (Zuk et al. 2014). Research has focused on the role and outcomes of male  
62 harmfulness on the persistence of polyandry, the effect of sperm competition on male  
63 ejaculate investment, the effects of kinship on male harmfulness, and a wide array of other  
64 topics (Rönn et al. 2006; Hotzy and Arnqvist 2009; Wilson and Tomkins 2015; Lymbery and  
65 Simmons 2017; McNamara et al. 2020). The aedeagus (intromittent organ) is covered in  
66 sclerotized spines that perforate the female reproductive tract during copulation, inflicting  
67 significant scarring (Crudginton and Siva-Jothy 2000; Dougherty and Simmons 2017). The  
68 degree of reproductive tract damage negatively impacts both female longevity and female  
69 reproductive success (den Hollander and Gwynne 2009) but promotes male fertilisation  
70 success (Hotzy and Arnqvist 2009). Reproductive tract damage facilitates the transport of  
71 accessory seminal compounds into the female bloodstream, which improve male competitive

72 fertilisation success (Hotzy et al. 2012; Rönn and Hotzy 2012; Yamane et al. 2015).  
73 Therefore, the harm to females in this species seems to be a side-effect of selection for  
74 competitive male fertilization success (Hotzy et al. 2012; McNamara et al. 2020). The  
75 thickness of the female reproductive tract wall appears to have evolved under sexually  
76 antagonistic coevolution to resist the harmful effects of male genital spines. Thus, across  
77 populations (Dougherty et al. 2017) and species (Ronn et al. 2007), female reproductive tract  
78 volume has been found to be positively correlated with male penile spine length, providing  
79 one of the few empirical examples of sexually antagonistic coevolution (Dougherty et al.  
80 2017). Moreover, male *C. maculatus* appear to adjust their copulatory behaviour (Wilson et  
81 al. 2014; Wilson and Tomkins 2014) and amount of harm imposed on females in response to  
82 their social environment [37,46, but see 47].

83         Here, we investigate whether female *C. maculatus* exposed to greater risk of sexual  
84 conflict can respond by adjusting the thickness of the reproductive tract to minimise  
85 anticipated male harm. We used a manipulation of the social environment to vary the  
86 immediate risk of sexual conflict: we had two treatments within which we manipulated both  
87 larval density, and the adult sex-ratio to simulate either a high or low sexual conflict risk.  
88 Additionally, we employed an experimental evolution design to test whether the thickness of  
89 the female reproductive tract diverged between populations of beetles evolving under a male-  
90 or female-biased sex-ratio. Assessment of reproductive tract morphology in this species has  
91 previously focused on either the thickness of the tract estimated from a small number of  
92 histological sections (Ronn et al. 2007), or the total volume of tissue across the entire tract via  
93 3-dimensional analysis of Micro-CT images (Dougherty et al. 2017). Although tissue volume  
94 may capture large-scale changes in female investment, it does not capture fine-scale changes  
95 in tract morphology. Here, we developed a novel technique to measure variation in the  
96 thickness of the female reproductive tract at different locations along its length using Micro-

97 CT data. We predicted that: 1) females in populations evolving under a male-biased sex-ratio  
98 will have thicker reproductive tract walls than those evolving under a female-biased sex ratio  
99 as a result of sexually antagonistic coevolution; and 2) females exposed to high-density larval  
100 environments and a male-biased social environment during development will develop thicker  
101 reproductive tract walls compared to females from low density larval environments exposed  
102 to a female-biased social environment, in anticipation of an increased risk of sexual conflict.

## 103 **2. Methods**

### 104 *(a) Study population*

105 The stock population of *C. maculatus* used for this study was derived originally from a  
106 population held by the CSIRO in 2005, which was itself founded by individuals found as  
107 agricultural pests. Both stock and experimental populations were maintained at 30°C under a  
108 12:12 hour day/night regime for the duration of the experiment (McNamara et al. 2016). For  
109 further information regarding the stock population see (Wilson et al. 2014).

### 110 *(b) Experimental evolution*

111 Experimental evolution lines were produced and maintained as described in McNamara *et al.*  
112 [18]. In brief, individuals from the stock population were used to create six experimental  
113 evolution lines. These lines were randomly assigned to one of two treatments, with either a  
114 male- or female-biased sex ratio. Each generation consisted of 120 individuals, with an 80:40  
115 male to female ratio for male-biased lines, and vice versa for female-biased lines. To control  
116 for potential differences in larval competition between treatments, female-biased populations  
117 received 200g of mung beans (*Vigna radiata*) for oviposition, while male-biased populations

118 received 100g. Each new generation was created by isolating 300 beans per population within  
119 ventilated 1.5mL Eppendorf tubes. After the required adults had emerged, sex-biased  
120 populations were again formed. This procedure was continued for 47 generations, after which  
121 populations were placed under common garden conditions for two generations with sex-ratio  
122 parity to remove the potential for any non-genetic parental effects. Following the second  
123 generation of common-garden breeding, beans were placed within ventilated Eppendorf  
124 tubes.

125 *(c) Social manipulation*

126 Isolated beans were assigned to one of two social manipulations designed to alter an  
127 individual's perception of future sexual conflict, either high-risk or low-risk. Evidence  
128 suggests that seed beetle larvae are able to determine population density before emergence  
129 from their beans via vibrations (Utida 1972; Thanthianga and Mitchell 1987). For this  
130 experiment we elected to use mung beans (*Vigna radiata*) as the larval host species to  
131 increase surface area compared to their larger, traditional host species (*Vigna unguiculata*),  
132 and thereby improve the transmission of vibrational cues to focal individuals. Therefore, in  
133 the high-risk treatment, five infested beans, each containing two larvae (infestation density  
134 can be assessed as eggs remain visible on the surface of the bean), were placed within an  
135 Eppendorf tube (resulting in ten larvae maximally per tube). Eppendorf tubes were checked  
136 daily for beetle emergence. Females that emerged synchronously with a male were discarded  
137 from the experiment to ensure focal females had standardised pre-mating social exposure and  
138 remained unmated. Females that emerged alone were placed within the lid of a 1.5mL  
139 Eppendorf tube, separated from four stock males and two stock females within the body of  
140 the tube via cotton mesh. The mesh allowed focal females to detect the presence of



141 individuals but prevented them from copulating with the males. Previous studies have shown  
142 that this methodology elicits phenotypic responses to the social environment in both sexes of  
143 *C. maculatus* (van Lieshout et al. 2014; Wilson and Tomkins 2014). Thus, high-risk females  
144 experienced high larval density and a male-biased sex ratio. For the low-risk treatment, a  
145 single infested bean, containing a single larva, was placed within an Eppendorf tube  
146 containing four un-infested beans. Following emergence, females were placed within an  
147 Eppendorf tube separated from two stock males and four stock females. Thus, low-risk  
148 females experienced a low larval density and a female-biased sex ratio prior to mating. In the  
149 low-risk social treatment, we aimed to provide cues that were representative of naturalistic  
150 conditions. Given the high densities experienced by *C. maculatus* when infesting food stores,  
151 we therefore exposed females to a small number of males rather than no males. For both  
152 treatments, females were removed after 24 hours and then allowed to mate once with a stock  
153 male. Females were isolated for a further 24 hours post-copulation, and then euthanized by  
154 freezing. Although we might expect adjustments in the reproductive tract to most likely occur  
155 during larval and pupal development, we also included the adult manipulation, because post-  
156 emergence sexual maturation is common in many insects, and this species has been found to  
157 respond to manipulations of sexual conflict at the adult stage (van Lieshout et al. 2014;  
158 Wilson and Tomkins 2014).

159

#### 160 *(d) Micro CT-scanning and tomographic reconstruction*

161 A total of five females per population-by-social-treatment combination were selected for  
162 Micro-CT scanning. Therefore, a total of 60 individuals were scanned for the purposes of this  
163 study. Sample tissue staining and scanning procedures followed those outlined by Dougherty

164 et al. [22], with the exception that we used formalin for tissue fixation rather than  
165 paraformaldehyde (see detailed pre-scan methodology in the online supplementary material).

166 Samples were scanned using a ZEISS Xradia Versa 520 X-ray microscope housed at  
167 the University of Western Australia Centre for Microscopy, Characterisation and Analysis.  
168 Samples were suspended in 100% ethanol and mounted in heat-sealed pipette tips with wax-  
169 covered tops in groups of 3-5 for scanning. Abdomens were arranged vertically and scanned  
170 sequentially from the highest abdomen to the lowest. Source voltage and power of initial  
171 scans (n = 11) was set at 40kV and 3W. However, source stability was compromised for an  
172 extended period of time, so the remaining scans were conducted at 60kV and 5W (n = 43) to  
173 ensure that scan quality remained stable (details of the scanning procedures can be found in  
174 the online supporting material). A total of six samples could not be analysed due to low scan  
175 quality, leaving a sample size of 54 for image analysis.

176

#### 177 *(e) Image analysis*

178 Images were analysed blind with regard to both the population of origin and social treatment.  
179 The analyses were performed using a combination of three custom-written FIJI (Schindelin et  
180 al. 2012; Schneider et al. 2012; Legland et al. 2016) scripts and Amira 6.2 (Thermo-Fisher  
181 Scientific, U.S.A.). We first selected a consistent region of interest within each reproductive  
182 tract, which was marked by the entrance of the spermathecal duct into the reproductive tract  
183 at one end (Figure 1a) and the first occurrence of bursal teeth on the other (Figure 1b) This  
184 region was chosen because it sustains the greatest damage during copulation (Dougherty et al.  
185 2017). Within this region of interest, we computed the 3D thickness of the dorsal and ventral  
186 reproductive tract walls based on the local thickness definition proposed by Dougherty and  
187 Kunzelmann (Dougherty and Kunzelmann 2007) (see online Supplementary material for

188 detailed methodology). According to this definition the thickness at any point within an  
189 object, in our case the tract walls, is the diameter of the largest sphere that fits inside the  
190 object and at the same time contains the point (Figure 1c). Further, we differentiated  
191 investment in the upper and lower reproductive tract by placing a horizontal plane running  
192 through the lumen's centroid, which allowed us to reliably define the upper and lower regions  
193 of tracts among all sampled individuals (Figure 1d).

194

#### 195 *(f) Statistical Analyses*

196 All statistical analyses were conducted in R (v 3.5.3) (R Development Core Team 2016).  
197 Normality of data was confirmed with Shapiro-Wilk's tests. Further, Bartlett's test for  
198 homogeneity of variance was found to be non-significant in all cases.

199 Pearson's correlation tests using the 'ppcor' R package (Kim 2015) found significant  
200 correlations between the mean, minimum and maximum thickness values from the upper and  
201 lower reproductive tract. Repeatability analyses were conducted for all measurements by  
202 extracting measures of 9 individuals on each of three separate occasions and analysing the  
203 data using the R package 'rptR' (Stoffel et al. 2017). We found significant repeatability ( $R >$   
204 0.6) in mean and maximum tract thickness but not in minimum tract thickness (see Table S2  
205 in the online Supplementary Material). We therefore conducted principal components  
206 analysis using the 'FactoMineR' R package, excluding upper and lower minimum thickness  
207 (Lê et al. 2008 Mar 18). One principal component (PC1) had an eigenvalue greater than 1 and  
208 was extracted for further analysis. PC1 was used as the dependent variable using a Gaussian  
209 linear mixed model that included female weight as a covariate. Following Arnqvist (Arnqvist  
210 2020), replicate population was included as a random factor within which an interaction  
211 between female weight and social treatment was fitted as a random slope. The random slope

212 allows for the interaction to vary across the differing populations (Arnqvist 2020). The source  
213 voltage was also included in this model as a random factor to control for any variance in trait  
214 estimates that might have arisen from the use of different voltage settings during scanning.  
215 The significance of our treatments, random factors and interaction effects were tested using  
216 Kenward-Roger F-tests using the R package ‘pbkrtest’(Halekoh and Højsgaard 2014). Non-  
217 significant interactions that did not improve model fit were subsequently removed from final  
218 models. Effect sizes were estimated as the standardised Pearson’s correlation coefficients  
219 (Table 3) using the ‘effectsize’ package in R. Estimated marginal means for each level of the  
220 two treatments were produced via the package ‘emmeans’. The data were explored for  
221 outliers using robust kernel-based outlier factor algorithms within the ‘OutlierDetection’ R  
222 package (k=3, bootstraps= 50,000). No outliers were identified.

### 223 **3. Results**

224 The first principal component explained 72% of the variance in reproductive tract thickness  
225 (Table 1) and was loaded equally by the mean and maximum thickness from both the upper  
226 and lower reproductive tract. There was no significant impact of evolutionary history or  
227 social treatment on female tract (Table 2). All interaction combinations between evolutionary  
228 history, social treatment and weight were found to be non-significant and were dropped from  
229 the final model.

230

231

### 232 **4. Discussion**

233 We developed a novel Micro-CT imaging method to measure the thickness of the  
234 reproductive tract walls of female *C. maculatus* seed beetles, from populations that had  
235 evolved under a male- or female-biased population sex ratio for 47 generations, and which  
236 were subsequently exposed to a social environment which conveyed either a high or low-risk  
237 of sexual conflict. Previous studies of this species have found that males can adjust their  
238 harmfulness to females in response to their social environment [43–45, but see: 56]. There  
239 was no effect of evolutionary history on the overall thickness of the female reproductive tract,  
240 nor was there any effect of the social environment. Our data therefore suggests that female  
241 reproductive tracts may not respond to variation in the risk of sexual conflict.

242         Previous studies using *C. maculatus* have similarly failed to find evolutionary  
243 responses in females to experimental manipulations of sexual conflict. Gay et al. [57] showed  
244 that after 90 generations of enforced monogamy, the re-introduction of polyandry over 30  
245 generations resulted in the evolution of harmful males but not resistant females. Similarly,  
246 after 8 generations of enforced monogamy it was found that the elaboration of male penile  
247 spines decreased, as might be predicted for a costly trait that functions in the context of  
248 competitive reproductive success. However, the morphology of the teeth found within the  
249 female genital tract failed to exhibit a correlated response to enforced monogamy (Cayetano  
250 et al. 2011). Our results are also consistent with recent findings of McNamara et al. [18] who  
251 utilised the same experimental evolution lines used in the current study. McNamara et al.  
252 (McNamara et al. 2020) found that although males evolving under a male-biased sex ratio  
253 evolved to be more harmful, females from populations evolving under a male-biased sex ratio  
254 experienced comparable reproductive tract scarring to females from populations evolving  
255 under a female-biased sex ratio. We found no evidence that sexual conflict intensity results in  
256 an evolutionary divergence in female reproductive tract thickness. Collectively, these  
257 findings suggest that either females fail to coevolve as readily as males, or that female

258 reproductive tract coevolution is more difficult to detect (Gay et al. 2010; Cayetano et al.  
259 2011; McNamara et al. 2020).

260         The findings of these experimental evolution studies are in contrast to comparative  
261 studies that have found evidence for sexually antagonistic coevolution between male  
262 harmfulness and female resistance to harm among *C. maculatus* populations and among  
263 *Callosobruchus* species more widely (Ronn et al. 2007; Dougherty et al. 2017). Further, the  
264 coevolution of resistance traits and male persistence has been identified in comparative  
265 studies of other arthropod taxa such as water striders (Heteroptera: Gerridae) (Arnqvist and  
266 Rowe 2002b) and diving beetles (Coleoptera: Dytiscidae) (Bergsten et al. 2008). The  
267 apparent inability of female seed beetle reproductive tracts to respond evolutionarily to  
268 relatively short-term manipulations in sexual conflict might be attributable to the difficulty in  
269 detecting sexually antagonistic coevolution at a particular point in time (Arnqvist and Rowe  
270 2005; Perry and Rowe 2012; Kokko and Jennions 2014). This suggestion could explain why  
271 females seemingly did not respond to our selection treatments, as it is clear that divergence in  
272 male harmfulness was present in these populations 15 generations earlier (McNamara et al.  
273 2020). We are unable to say whether females from populations experiencing differing levels  
274 of sexual conflict might have developed alternative methods of resistance, such as higher  
275 immunocompetence (Hangartner et al. 2015). Previous studies utilising the same  
276 experimental evolution lines have shown that individuals derived from the male-biased lines  
277 exhibit reduced immune function (van Lieshout et al. 2014). These results are indicative of a  
278 resource trade-off between immune function and reproductive investment fuelled by the costs  
279 of high intensity sexual conflict. Although the method by which Gay et al. [57] and  
280 McNamara et al. (McNamara et al. 2020) altered sexual conflict intensity differed, both  
281 studies found that females from populations that experienced higher conflict were better able  
282 to counter-act the negative impact of mating on their fitness. However, it is clear that the

283 method by which female *C. maculatus* accomplish this is not via genital morphology, female  
284 kicking behaviour, or through improved immunity (van Lieshout et al. 2014; McNamara et al.  
285 2020). Finally, it may be that inbreeding depression was responsible for impeding divergence  
286 among our lines. This seems unlikely however, because a hallmark of inbreeding is reduced  
287 fitness, which would have been observed as reduced fecundity for females from the female-  
288 biased lines. A previous study using the same experimental evolution lines (McNamara et al.  
289 2020) showed no evidence that females from female-biased lines experienced reduce fitness  
290 compared to those from male-biased lines.

291 We found that the social environment had no impact on female reproductive tract  
292 thickness. This is the only study to have investigated whether females are able to plastically  
293 respond to sexual conflict risk by altering reproductive tissue dimensions. There are several  
294 reasons why females might have failed to respond to our manipulation of their social  
295 environment. First, it is possible that females were unable to adjust the reproductive tract in  
296 response to our environmental cues. Females were given a proxy for sexual conflict risk via  
297 manipulations of larval density during development, but this cue may have been insufficient  
298 to provoke a plastic response. Second, the larval and pupal stages are critical to the  
299 development of insect reproductive organs (Happ 1992). This is demonstrated by the long-  
300 lasting impacts of larval food availability on adult reproductive output in both female and  
301 male insects (Bauerfeind and Fischer 2005; Engels and Sauer 2007). Females were exposed  
302 to direct signals of increased sexual conflict risk (via sex-ratio) in their adult stage, post-  
303 pupation, but they may be unable to make plastic adjustments after adult emergence. Third, it  
304 is also possible that our measures of tract thickness do not capture important qualitative  
305 variation. For example, it may be that females can also plastically adjust the elasticity of the  
306 reproductive tract through the incorporation of resilin, a compound shown to improve  
307 tolerance to cuticle perforation in bed bugs (*Cimex lectularius*)(Michels et al. 2015). Finally,

308 it may be that females plastically responded to our social manipulation, but due to the  
309 logistical limitations placed on us with respect to the numbers of females we could scan, we  
310 were unable to capture any differences among them. However, given that our repeatability  
311 analysis revealed significant variation among females in reproductive tract thickness, we can  
312 be certain that we are able to detect variation in tract thickness using this novel technique. If  
313 there were an effect of the social environment on the reproductive tract thickness, the effect  
314 size estimated in our analysis suggest that it is small and would require a large sample size in  
315 order to detect significance.

316         Although our measure of female resistance traits is limited to a quantitative measure  
317 of reproductive tract thickness, it nonetheless offers a significant advance over previous  
318 studies. Previous studies utilising 3-dimensional analysis of Micro-CT scans have measured  
319 total reproductive tract volume, which is effective at controlling for tract shape and size  
320 effects, but cannot identify fine-scale changes in morphology within the reproductive tract  
321 (Dougherty et al. 2017). Our current method overcomes this challenge by allowing us to  
322 identify variation in thickness in the upper and lower regions of the reproductive tract in 3-  
323 dimensional space. Overall, our novel method for measuring reproductive tract thickness  
324 shows promise in its ability to detect fine-scale differences in internal structures of the female  
325 reproductive tract, and promises to be a valuable tool in the long-awaited study of female  
326 genital morphology across numerous species.

327         In conclusion, we provide no evidence for plastic adjustments of reproductive tract  
328 thickness, a trait known to have coevolved with male-imposed genital damage among  
329 populations and species of *Callosobruchus*. Moreover, we found that populations that had  
330 evolved under intense sexual conflict failed to diverge in female genital morphology. The  
331 coevolution of male and female genital traits among populations suggests ample genetic  
332 variation in tract morphology may exist, however among-population genetic variance does



333 not necessarily reflect within-population variation (Hoffmann et al. 2003). Therefore, the lack  
334 of divergence in tract morphology across our evolution lines may reflect a lack of within-  
335 population genetic variation for this trait. Studies of sexual selection acting on female  
336 genitalia typically lag behind those focussed on male traits (Kokko and Jennions 2014; Sloan  
337 and Simmons 2019). Investigating female traits and their responses to male harmfulness can  
338 broaden our understanding of sexual selection and sexual conflict. Further research needs to  
339 focus on the female perspective if we are to quantify the pervasiveness and intensity of  
340 female responses to sexual conflict. The barriers to such research are slowly dissipating with  
341 the advent of innovative new technologies, such as Micro-CT scanning, that allow more  
342 effective measurement of female traits. As technologies become more accessible and cheaper  
343 to employ, increased sample sizes will be possible so that future studies have the power to  
344 detect variation in these minute structures. We believe that the present study provides a viable  
345 methodological approach for further investigations into plastic adjustments in female  
346 reproductive tracts, which is flexible enough to identify small-scale variation.

347

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359

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546

547 **Table 1** Fit and loadings for the first two principal components explaining variation in female  
 548 reproductive tract thickness.

	<b>PC1</b>	<b>PC2</b>
Eigenvalue	<b>2.88</b>	0.78
% Variance	71.91	19.91
Upper mean	0.930	0.25
Upper maximum	0.777	0.60
Lower mean	0.877	-0.36
Lower maximum	0.811	-0.47

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550

551 **Table 2** Analysis of variance for fixed and random effects on a multivariate measure of  
 552 female reproductive tract thickness (PC1) (N=54).

	<b>F</b>	<b>df</b>	<b>p</b>	<b>Variance</b>
Evolutionary History	0.51	1, 3.79	0.52	
Social Environment (SE)	1.18	1, 4.38	0.34	
Body Weight	3.95	1, 3.61	0.12	
Population replicate				4.31
Source voltage				<0.01

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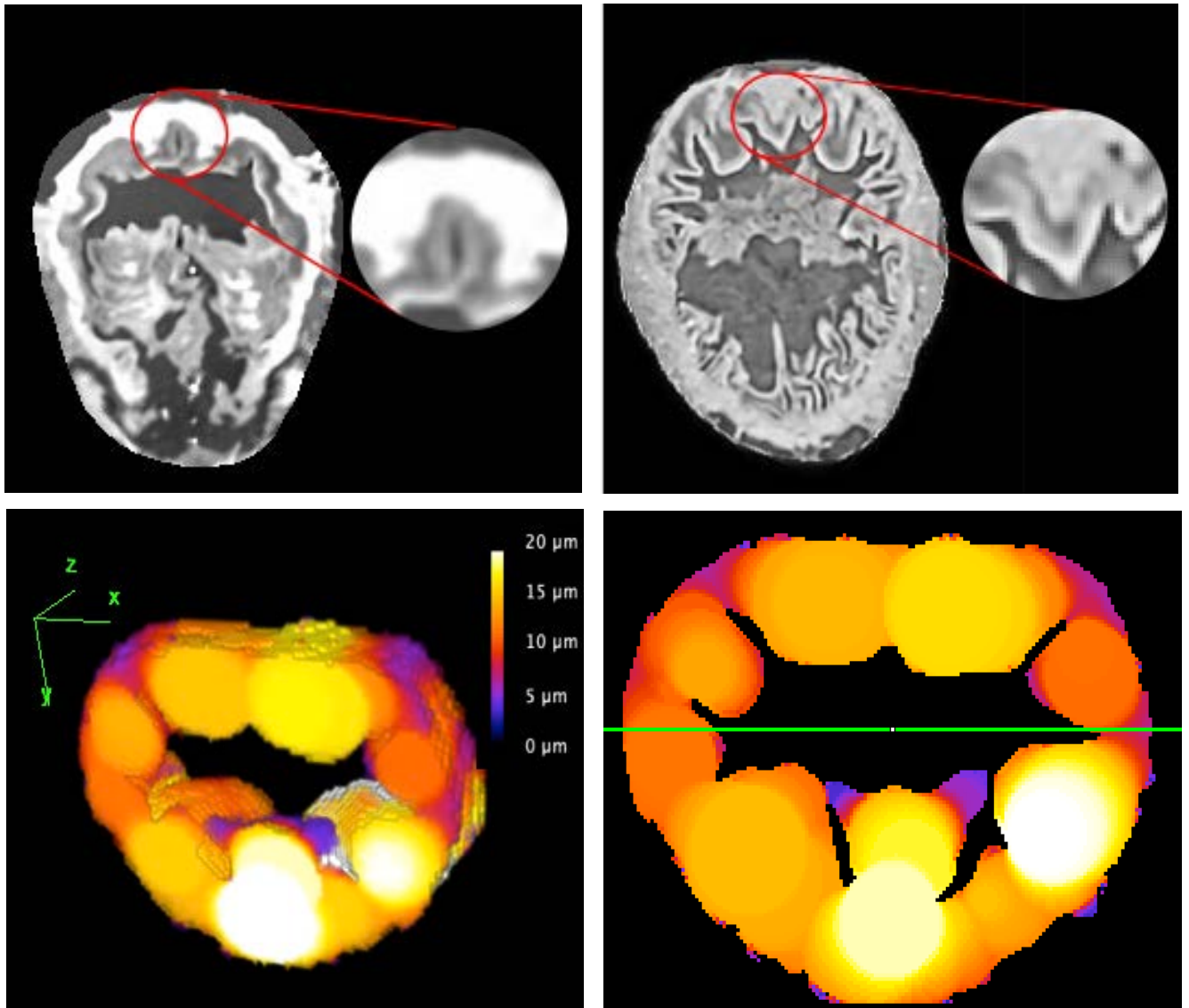
561

562 **Table 3** Estimated marginal means and effect sizes (Pearson's  $r$ ) with 95% confidence  
 563 intervals of all fixed effects on a multivariate measure of female reproductive tract thickness  
 564 (PC1) (N=54).

	Mean	95% CI	Effect size ( $r$ )	95% CI
<i>Evolutionary History (M-F)</i>			-0.28	-0.76, 0.21
Female-bias	0.638	0.013, 1.26		
Male-bias	0.580	-0.17, 1.33		
<i>Social Environment (L-H)</i>			-0.43	-0.92, 0.05
High-risk	0.643	0.047, 1.24		
Low-risk	0.575	-0.15, 1.30		

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568 **Figure 1** Segmentation of the reproductive tract: (a) First appearance of the entrance of the  
 569 spermathecal duct. (b) First appearance of the bursal teeth. Tract wall thickness: (c) 3D  
 570 thickness heat map (d) Upper and lower regions of interest within the reproductive tract.

571